**The following (Appendix 9 documents) were taken (verbatim) from ACORPs that were submitted to JIT for review. These examples will be part of the interactive session presented by Dr. Richerson, please review this information prior to the VA luncheon meeting scheduled for 12:00-2:00 pm Mountain Time on Tuesday, 11/3/15.**

**Example #1**

**ACORP Appendix 9**

**DEPARTURES FROM "MUST" AND "SHOULD" STANDARDS IN THE *GUIDE (2011)***

**VERSION 4**

See ACORP App. 9 Instructions, for more detailed explanations of the information requested.

For each IACUC-approved "departure" of this protocol from a "Must" or "Should" standard in the *Guide,* provide the following information. (Consult the IACUC or the Attending Veterinarian for help in determining whether any "departures" are involved.):

Copy the lines below for each departure.

Briefly summarize the "Must" or "Should" standard, and provide the number(s) of the page(s) on which it appears in the *Guide.*

**►Social animals should be housed in pairs or groups. Chapter 3, Page 51.**

Describe the specific alternate standard(s) that will be met on this protocol. and how they will be monitored .

**►Mice will be single housed: males after breeding. any animal fighting, those in nesting studies, those in running wheel studies.**

Provide the scientific, veterinary medical or animal welfare considerations that justify this departure

**►After having been single housed, male mice routinely fight when social housing is attempted. Single housing is required during running wheel studies because data is collected from a single animal; the instrument cannot distinguish between animals. Nesting study mice are only singly to score next building; they are housed very briefly (less than 24 h). Nesting animals may be rehoused with cagemates when testing is completed.**

**Example #2**

**ACORP Appendix 9**

**DEPARTURES FROM “MUST” AND “SHOULD” STANDARDS IN THE *GUIDE (2011)***

For each IACUC-approved “departure” of this protocol from a “Must” or “Should” standard in the *Guide*, provide the following information. (Consult the IACUC or the Attending Veterinarian for help in determining whether any “departures” are involved.):

Copy the lines below for each departure.

Briefly summarize the “Must” or “Should” standard, and provide the number(s) of the page(s) on which it appears in the *Guide.*

**► Euthanasia by Decapitation without anaesthesia:**

**AVMA Guidelines (2013) Page 61, Section S2.2.2.3: Small Lab Animals (Rats), Acceptable with Conditions Methods, Physical methods, Decapitation**

**“Decapitation is used in lab settings because it yields tissues uncontaminated by chemical agents. Loss of cortical function is rapid and occurs within 5-30 seconds. ”**

Describe the specific alternate standard(s) that will be met on this protocol, and how they will be monitored.

► **Decapitation is acceptable with conditions for rodents:**

**1. Use of a specialized rat guillotine, maintained in good condition with sharp blades.**

**2. Personnel will be or have been trained on anaesthetized and/or dead animals to demonstrate proficiency. Only trained personnel are allowed to euthanize animals on this protocol.**

**3. We will use restraints (Decapicones, transparent plastic cones open at the end) to prevent excess moving around of rats just prior to decapitation, which allows the procedure to be done quickly and reduce unnecessary distress on the animals.**

Provide the scientific, veterinary medical, or animal welfare considerations that justify this departure

**►In this study, we plan to determine post-mortem levels of monoamines (serotonin and dopamine) and monoamine receptor levels in brain regions involved in depression. Inhalant agents and barbiturates are documented to alter monoamine transporter function (el-Maghrabi and Eckenhoff., 1993; Fink-Jensen et al., 1994; Mantz et al., 1994) and would prevent accurate measurement of rat brain monoamines in our study. A physical method of euthanasia without sedation is therefore necessary in this study.**

**Description: Decapicones (tapered plastic tubes, Braintree Scientific Inc.) will be used to restrain rats prior to live decapitation using a guillotine, a process that is quick and relatively stress-free for the animals. The guillotine is cleaned with soap and water immediately after each use and rinsed with 70% ethanol prior to use again. The guillotine is maintained with a sharp blade to provide instant decapitation.**

• **El-Maghrabi, EA, and RG Eckenhoff (1993) Inhibition of dopaminergic transport in rat brain synaptosomes by volatile anesthetics. Anesthesiology 78: 750.**

• **Fink-Jensen A, SH Ingwersen, PG Nielsen, L Hansen, EB Neilsen and AJ Hansen (1994) Halothane anesthesia enhances the effect of dopamine uptake inhibition on interstitial levels of striatal dopamine. N. Sch. Arch. Pharmacol. 350:239.**

• **Mantz, J., C Varlet, JB Lecharny, D Henzel, P Lenot and JM Desmonts (1994) Effects of volatile anesthetics, thiopental and ketamine on spontaneous and depolarization-evoked dopamine release from striatal synaptosomes in the rat. Anesthesiology 80: 352.**

**Example #3**

**ACORP Appendix 9**

**DEPARTURES FROM "MUST" AND "SHOULD" STANDARDS IN THE *GUIDE (2011)***

**VERSION 4**

See ACORP App. 9 Instructions, for more detailed explanations of the information requested.

For each IACUC-approved "departure" of this protocol from a "Must" or "Should" standard in the *Guide,* provide the following information. (Consult the IACUC or the Attending Veterinarian for help in determining whether any "departures" are involved.):

Copy the lines below for each departure.

Briefly summarize the "Must" or "Should" standard, and provide the number(s) of the page(s) on which it appears in the *Guide.*

**►The recommended minimum space for commonly used laboratory rodents housed in groups of a female Dam + Litter is 51 in2 of floor area. Mice >25g require >15 in2 each. Sufficient space should be allocated for mothers with litters to allow the pups to develop to weaning without detrimental effect to the mother or the litter.**

Describe the specific alternate standard(s) that will be met on this protocol, and how they will be monitored.

**►**

1. **The two females should give birth no more than four days apart. If the second female has not given birth after four days, she should be moved to a new cage.**
2. **Strains and stocks that routinely have large litters (avg. 10+ pups) may not be bred in trios.**
3. **Restrictions on #pups/cage are imposed when trio breeding.**
4. **The VMU Supervisory staff and Veterinary staff will have the discretion to request animals to be separated to new cages if determined that the home cage is overcrowded, and/or the animal welfare is of concern.**

Provide the scientific, veterinary medical, or animal welfare considerations that justify this departure.

**►The maximum number of pups in one cage is no more than 12: if greater, litter size will be culled**

 **to 12 by P12 of the youngest litter if 2 litters are present.**

**VA Deviations and Departures Flowchart**

